

# INTERNATIONAL SYMPOSIUM POTABLE REUSE

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## The limitations of common molecular techniques for water reuse microbiology

Rose Kantor, Ph.D., Scott Miller, Lauren Kennedy, and Prof. Kara Nelson  
University of California, Berkeley

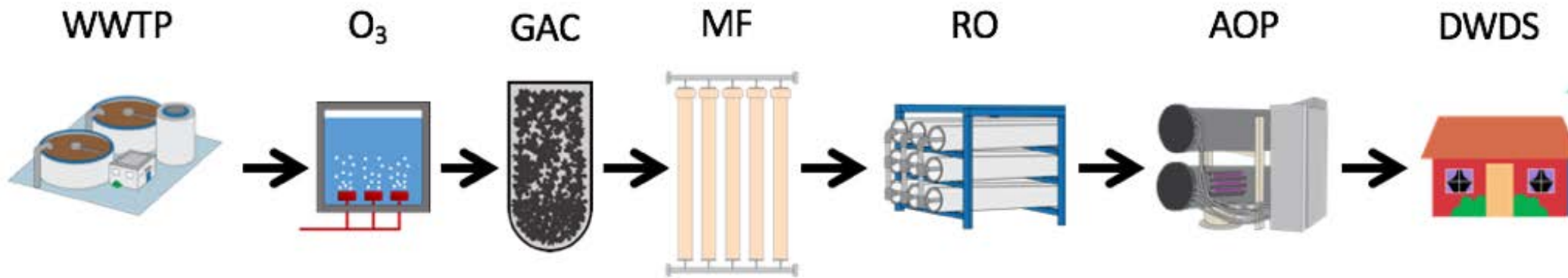


**American Water Works  
Association**

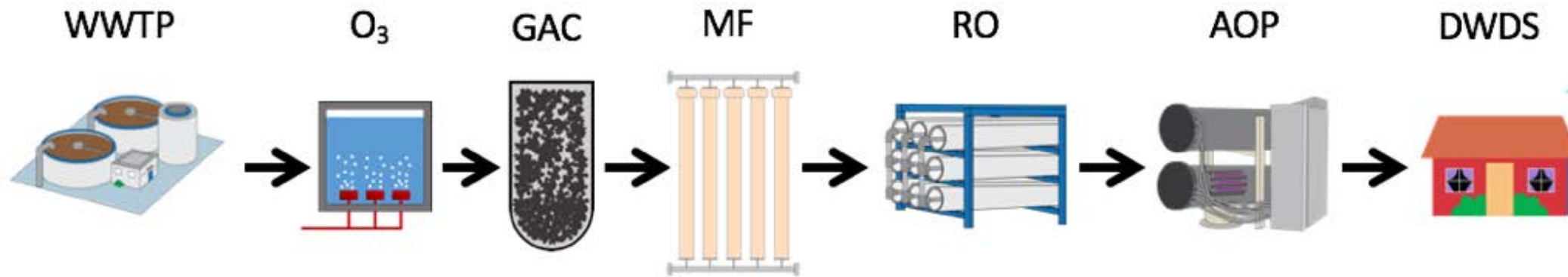
# Talk outline

1. Motivation: why do we care about bacteria?
2. Methods: what tools are available?
3. Amplicon sequencing: the juicy details
4. Results from sequencing at advanced treatment pilot
5. Complementary methods

# Motivation: potable reuse microbiology

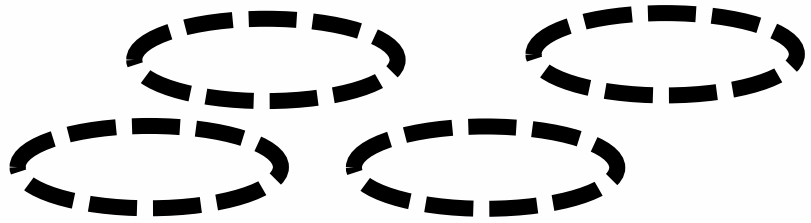


# Motivation: potable reuse microbiology

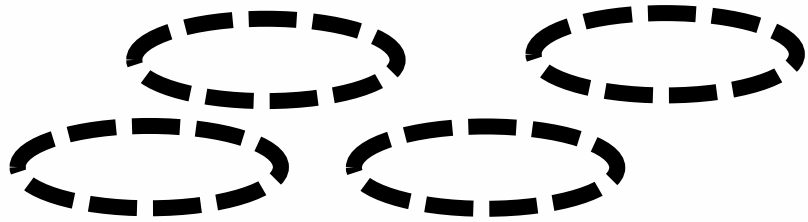


- What types of microorganisms are present and in what quantities?
- Where might the bacteria come from? (source tracking)

## Non-molecular methods

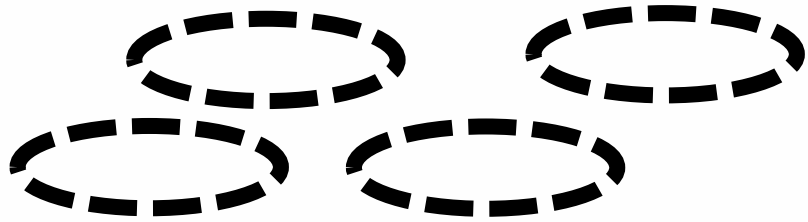


## Non-molecular methods



- Total coliforms
- Heterotrophic plate count
- ATP
- Flow cytometry

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## Molecular methods



- PCR and qPCR
- 16S rRNA gene amplicon sequencing
- Metagenomics (shotgun sequencing)

## Molecular methods



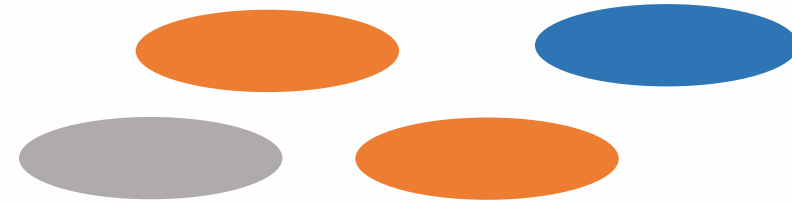
*Targeted quantification*  
Low quantities of DNA



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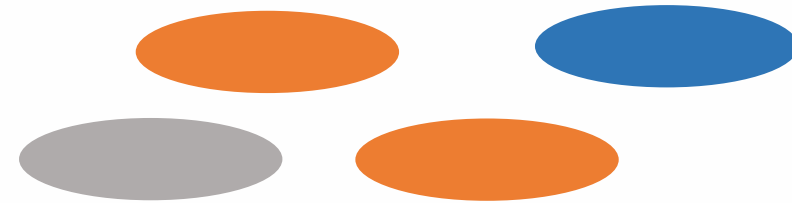
*Broadest identification*

Higher quantities of DNA



- Metagenomics (shotgun sequencing)

Molecular methods



*Targeted quantification*

Low quantities of DNA



• PCR and qPCR

*Broad identification*

Low quantities of DNA



• **16S rRNA gene amplicon sequencing**

*Broadest identification*

Higher quantities of DNA



• Metagenomics (shotgun sequencing)

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?

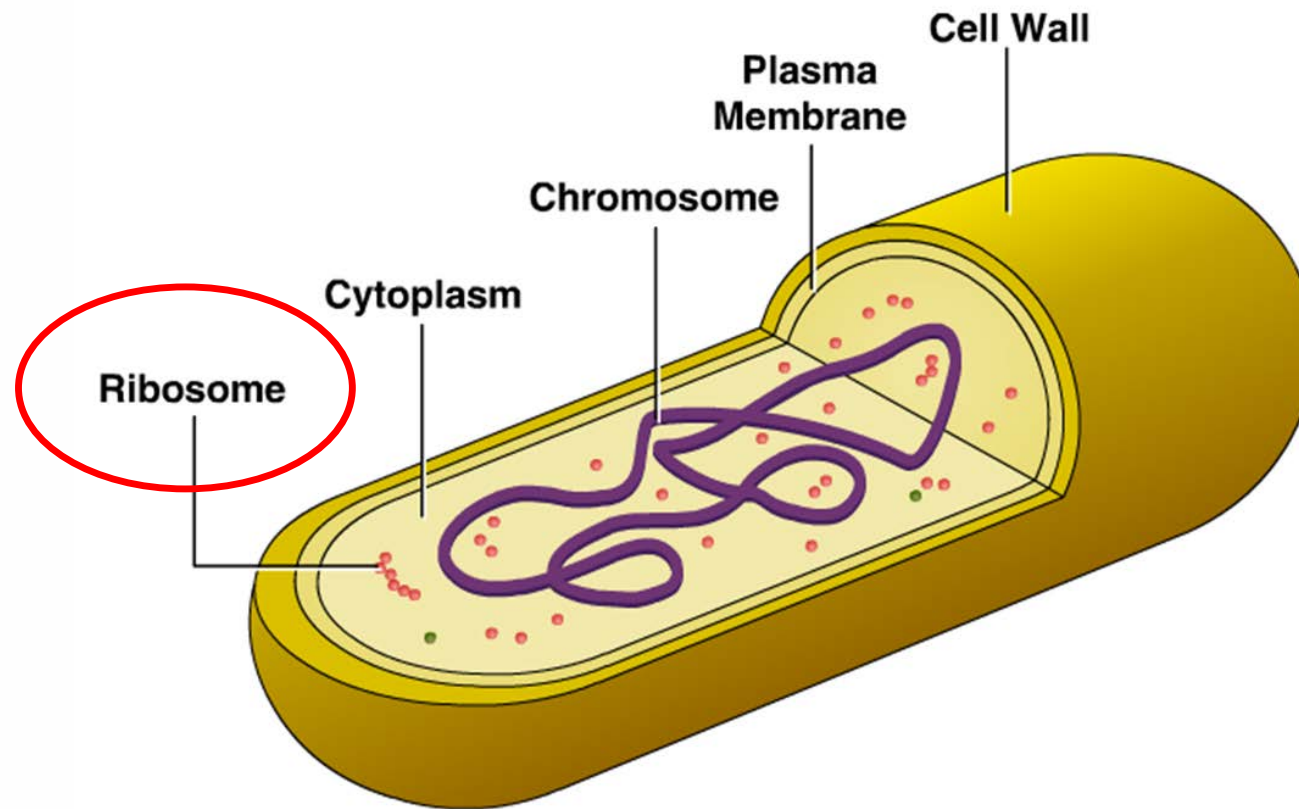
# Amplicon sequencing is like bug collecting



# DNA sequences can be used for identification

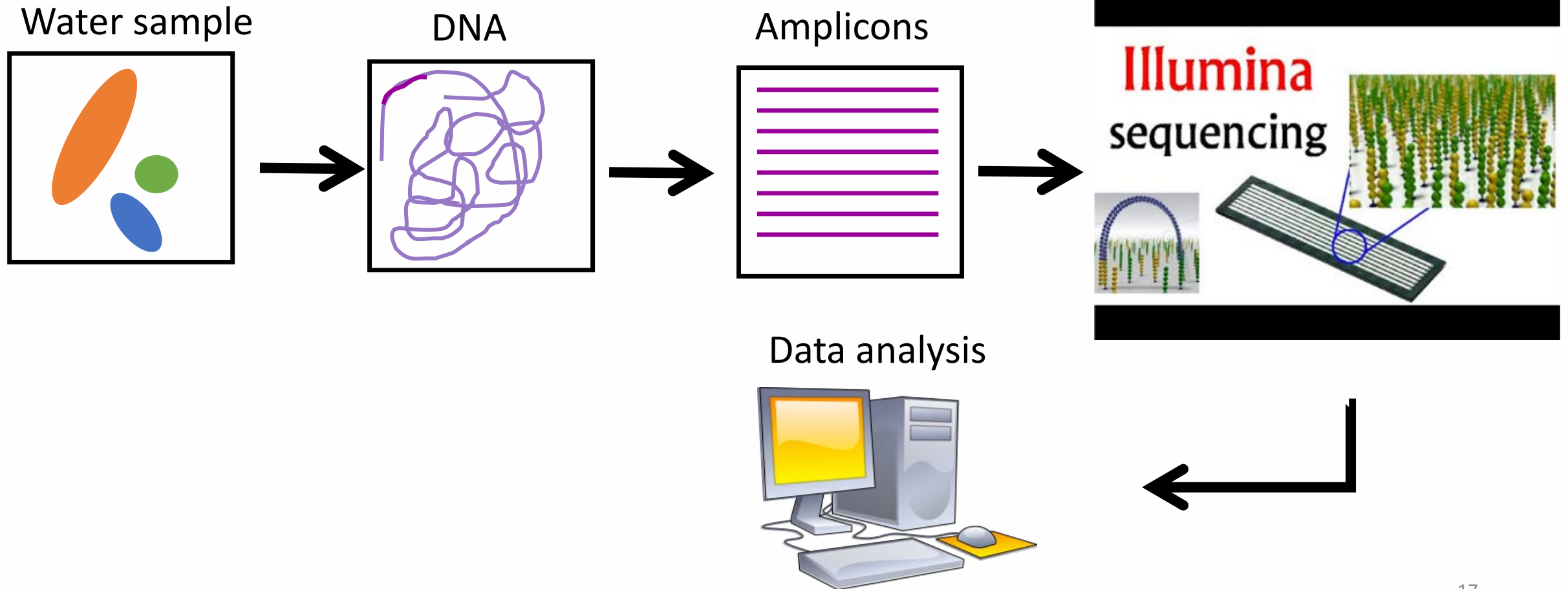
	1	10	20	30	40	50	60																																																								
1. seq1	C	G	T	T	G	C	T	C	G	G	A	A	T	C	A	C	T	G	G	G	C	G	T	A	A	A	G	G	C	G	C	G	T	A	G	G	C	G	G	C	G	T	T	T	T	A	A	G	T	C	G	G	G	G	G	T	G	A	A	A	G	C	
2. seq2	C	G	T	T	G	T	C	C	G	G	A	A	T	T	A	C	T	G	G	G	C	G	T	A	A	A	G	A	G	C	T	C	G	T	A	G	G	T	G	G	T	T	G	T	C	G	C	G	T	T	G	T	T	C	G	T	G	A	A	A	A	C	
3. seq3	C	G	T	T	G	T	C	C	G	G	A	A	T	T	A	C	T	G	G	G	C	G	T	A	A	A	G	A	G	C	T	C	G	T	A	G	G	T	G	G	T	T	G	T	C	A	C	G	T	T	G	T	C	C	G	T	G	A	A	A	A	C	
4. seq4	C	G	T	T	G	C	T	C	G	G	A	A	T	C	A	C	T	G	G	G	C	G	T	A	A	A	G	G	G	C	G	C	G	T	A	G	G	C	G	G	T	T	T	T	A	A	G	T	C	G	G	G	G	G	T	G	A	A	A	G	C		
5. seq5	C	G	T	T	G	T	C	C	G	G	A	T	T	T	A	T	T	G	G	G	C	G	T	A	A	A	G	A	G	T	T	C	G	T	A	G	G	C	G	G	T	T	G	T	T	A	A	G	T	C	T	G	A	T	G	T	T	A	A	A	G	A	
6. seq6	C	G	T	T	G	C	T	C	G	G	A	A	T	C	A	C	T	G	G	G	C	G	T	A	A	A	G	G	G	T	G	C	G	T	A	G	G	C	G	G	G	T	C	T	T	T	A	A	G	T	C	A	G	G	G	G	T	G	A	A	A	T	C

Amplicon sequencing is based on ribosomal RNA genes

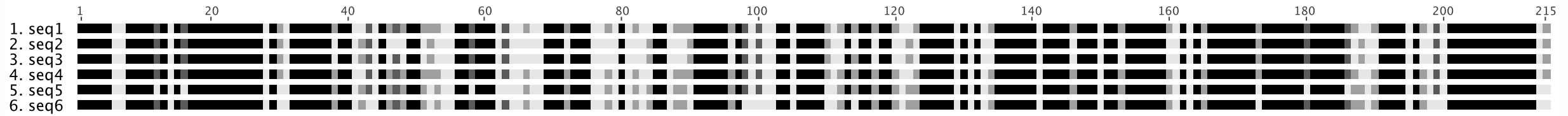




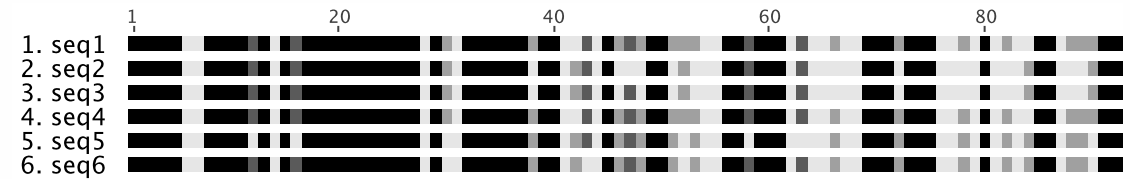
# Amplicon sequencing process



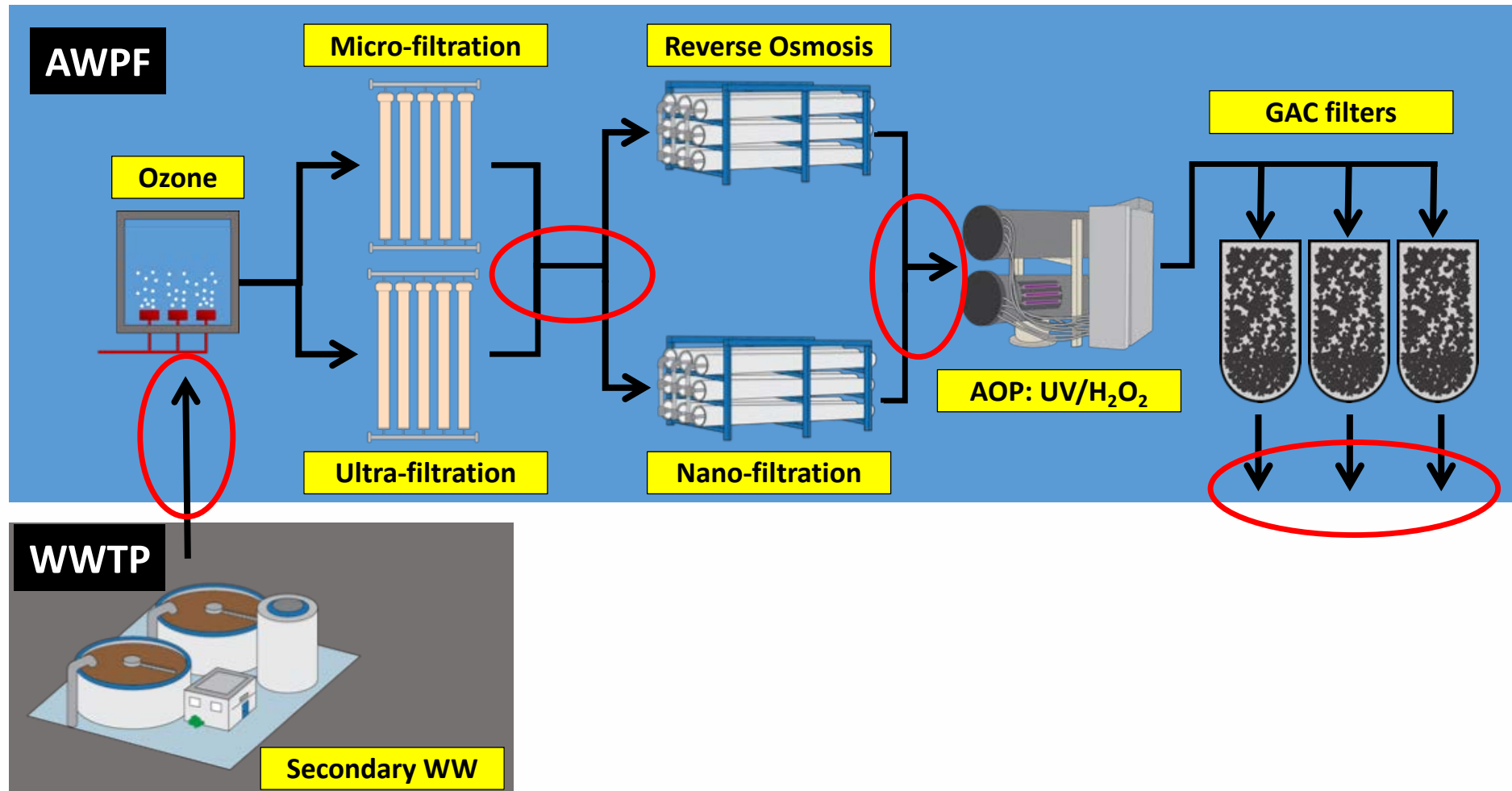
# 16S rRNA genes show relatedness of bacteria



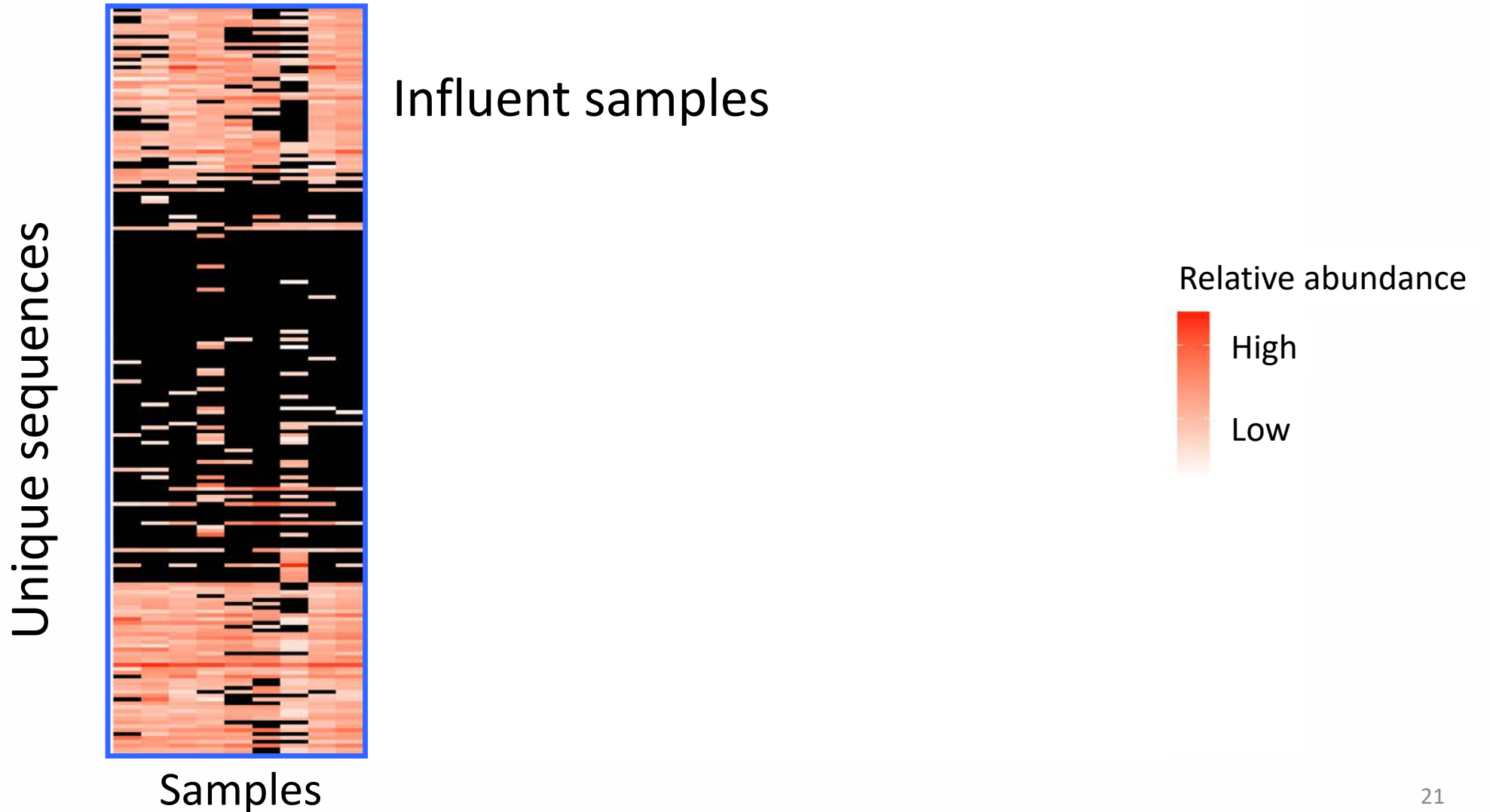
# Identification via amplicon sequencing



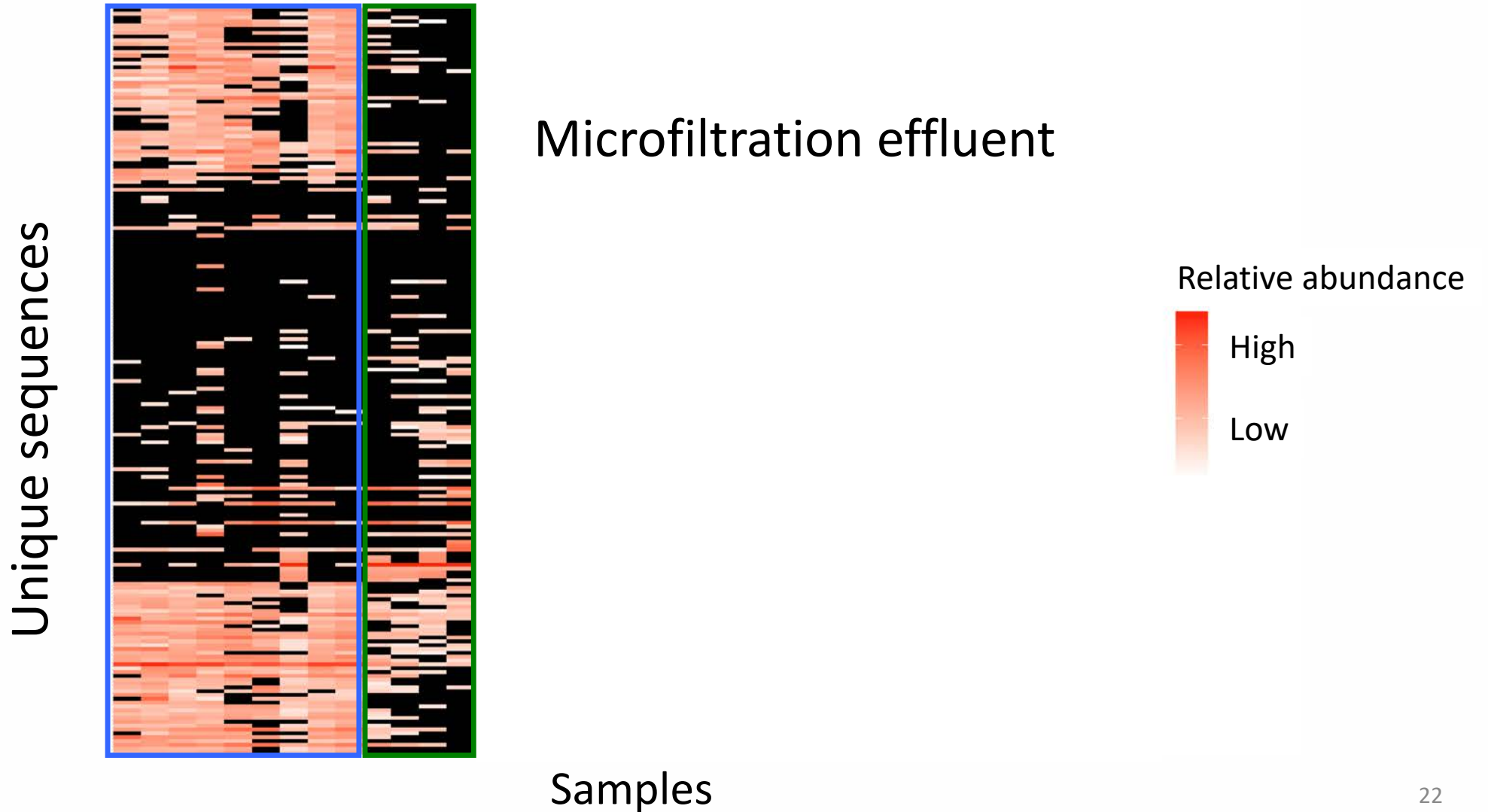
# Amplicon seq. on El Paso DPR treatment train



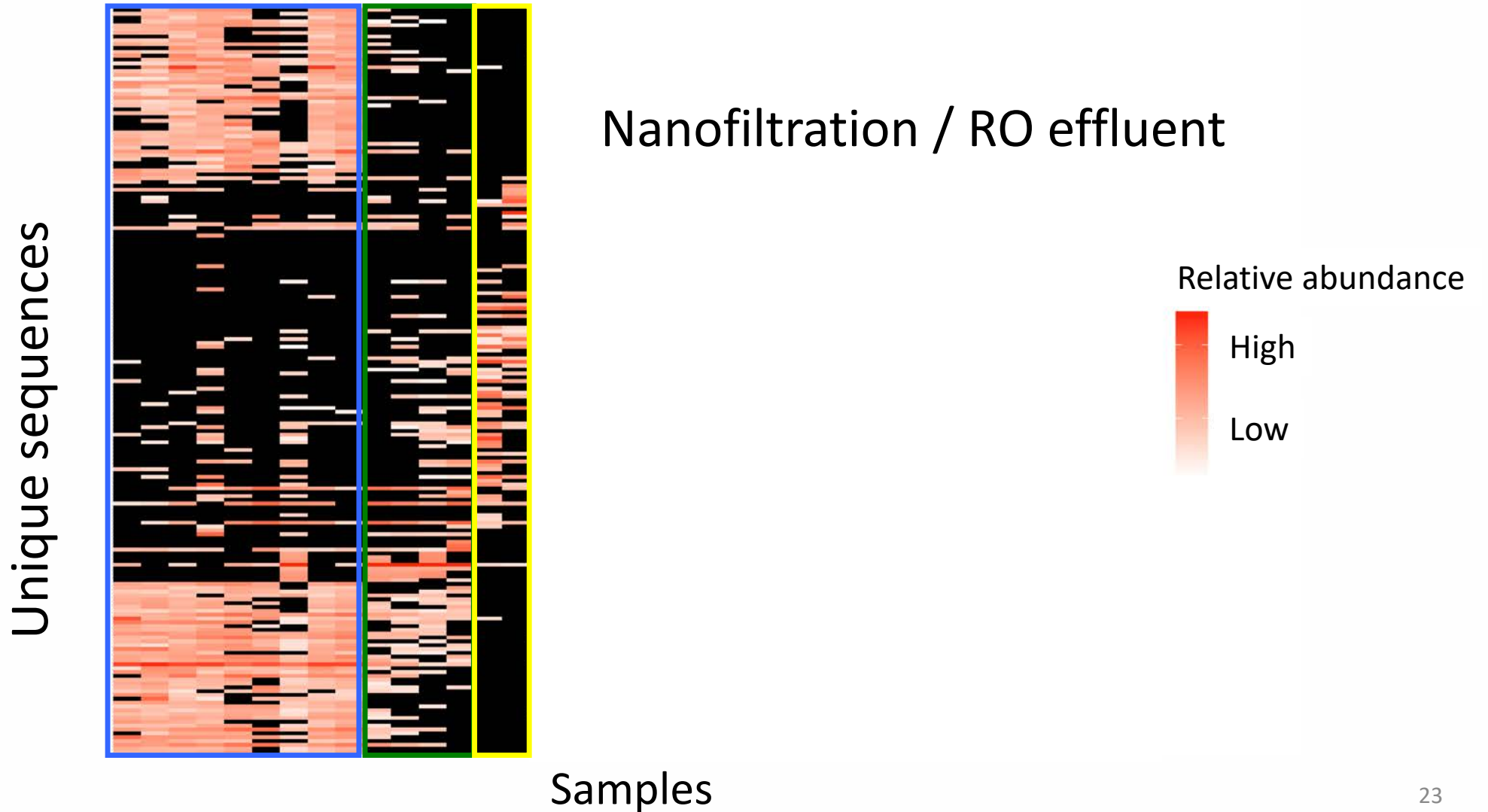
# Results: amplicon seq. across treatment train



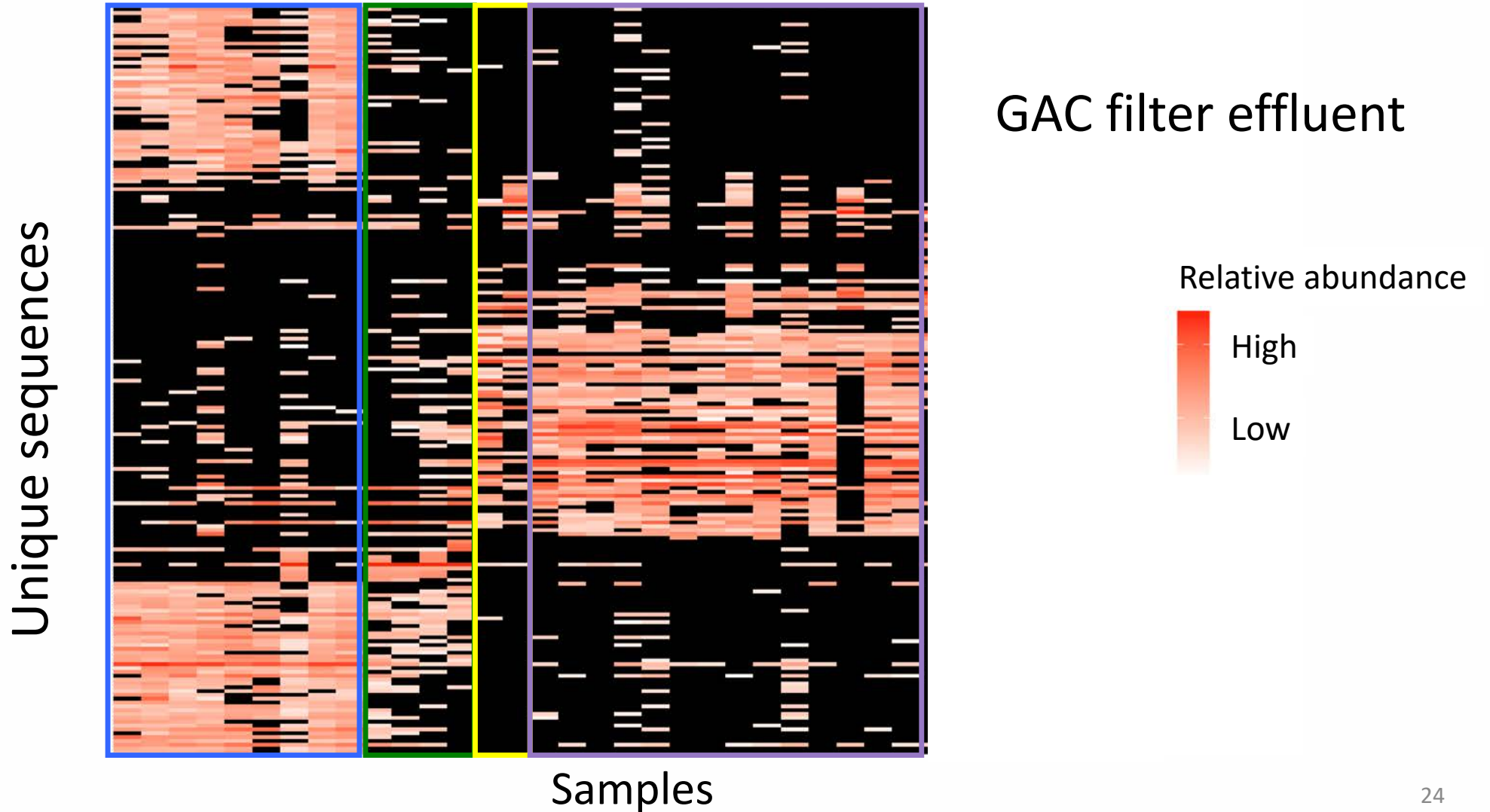
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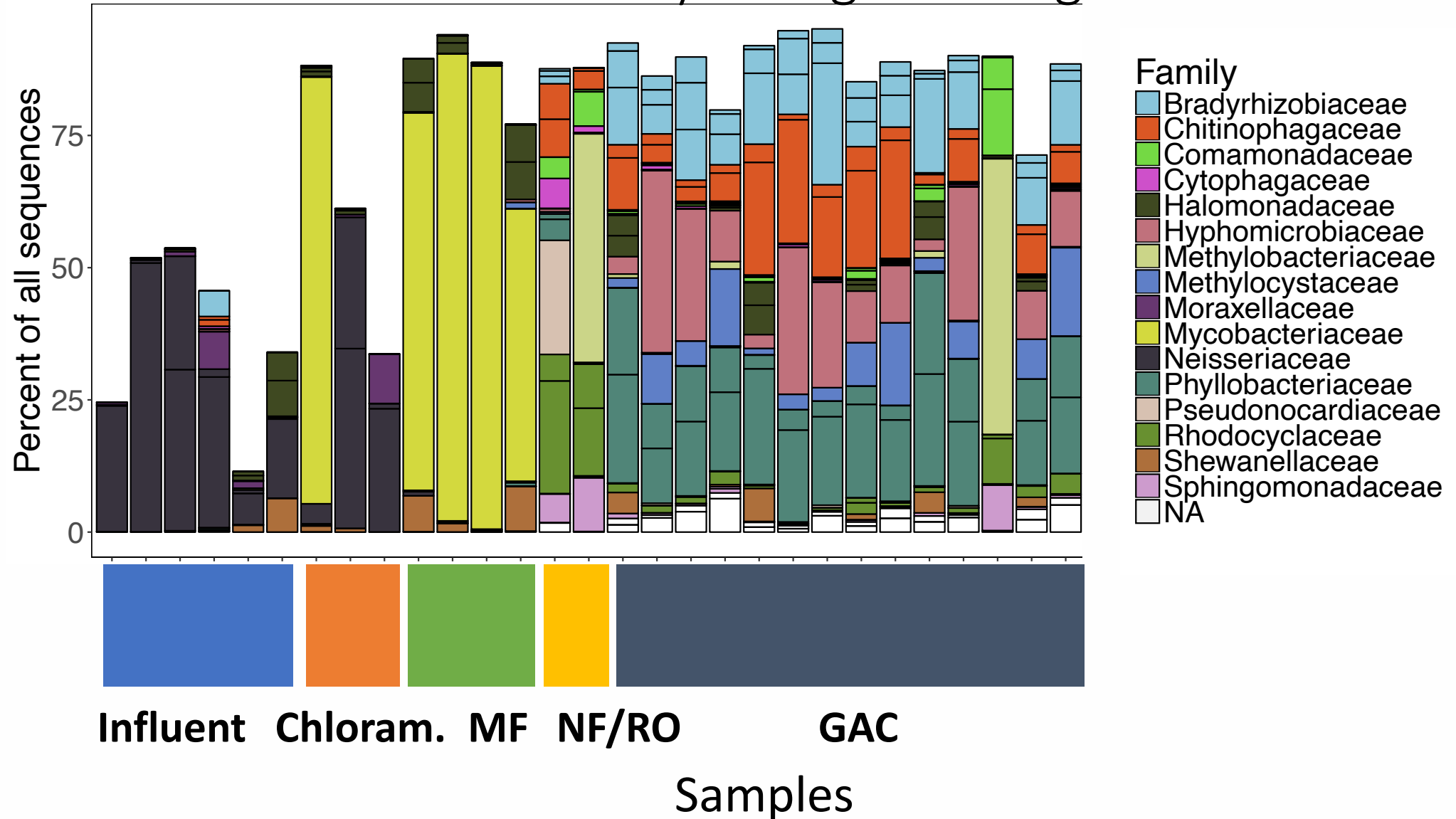


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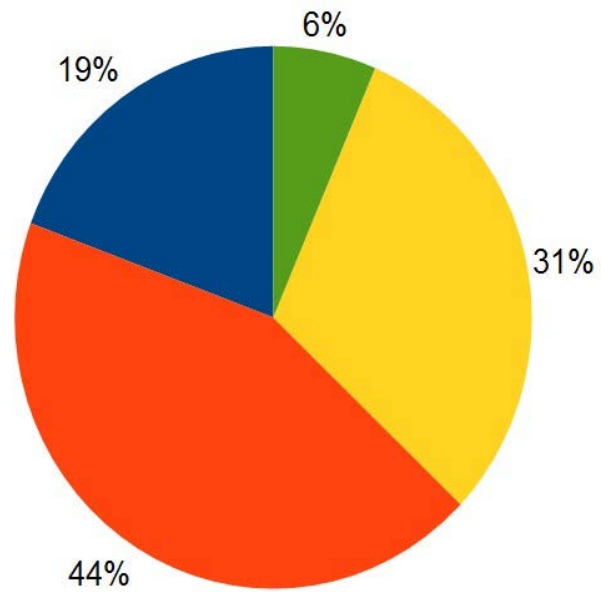




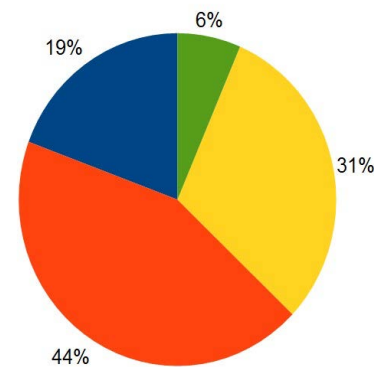
# Results: microbial community changes during treatment



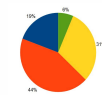
# Limitation 1: Relative vs. Absolute Abundance



Influent



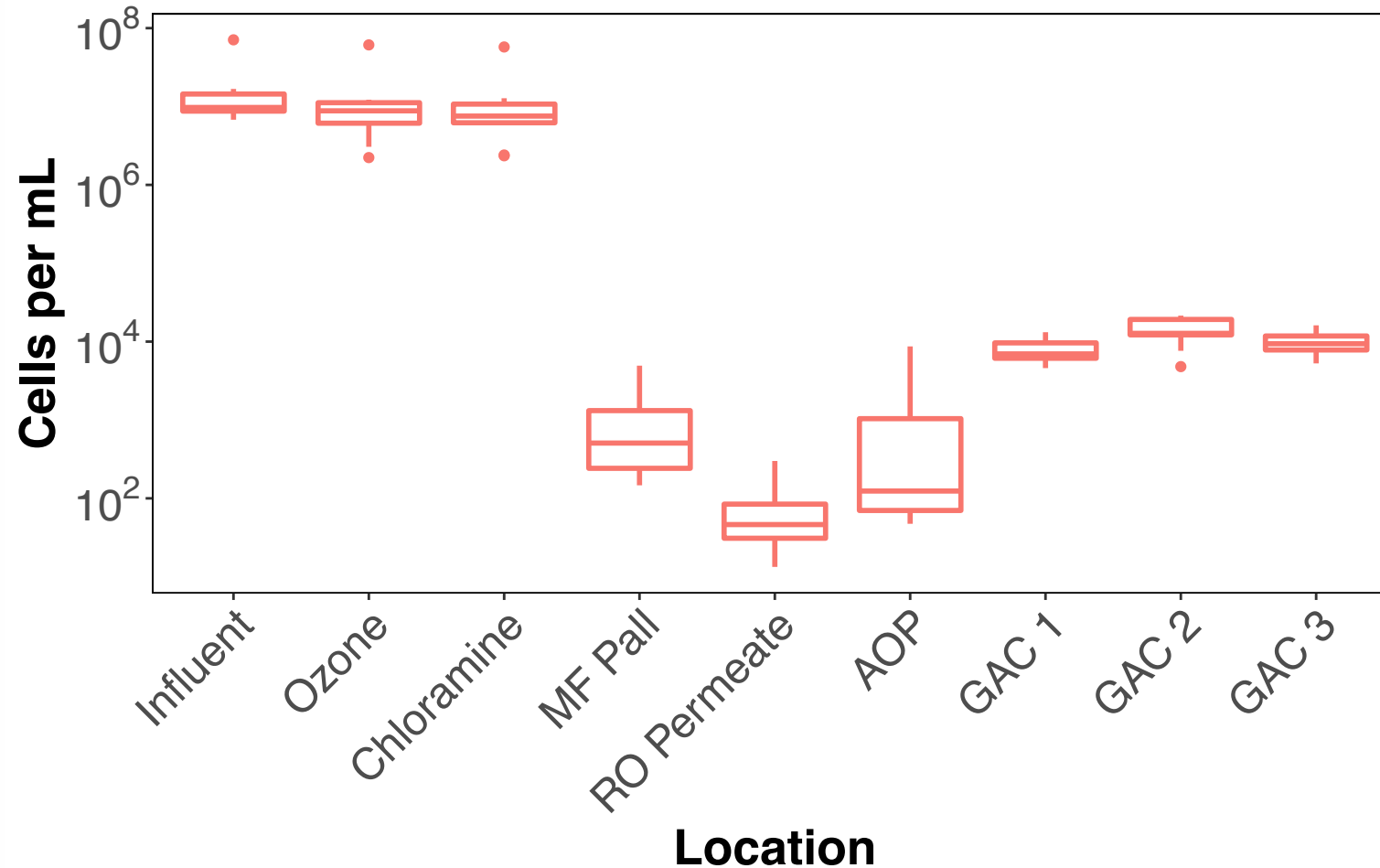
MF effluent



RO effluent

# Absolute quantification

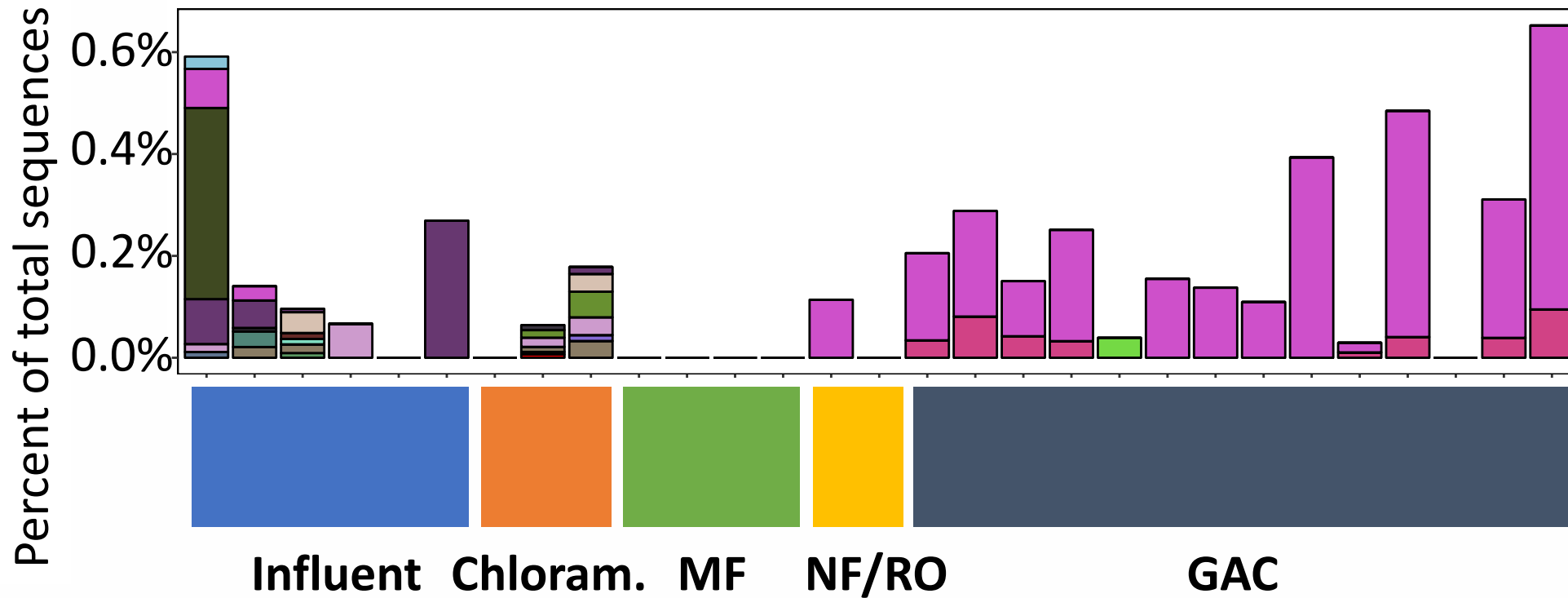
Cell counts via flow cytometry



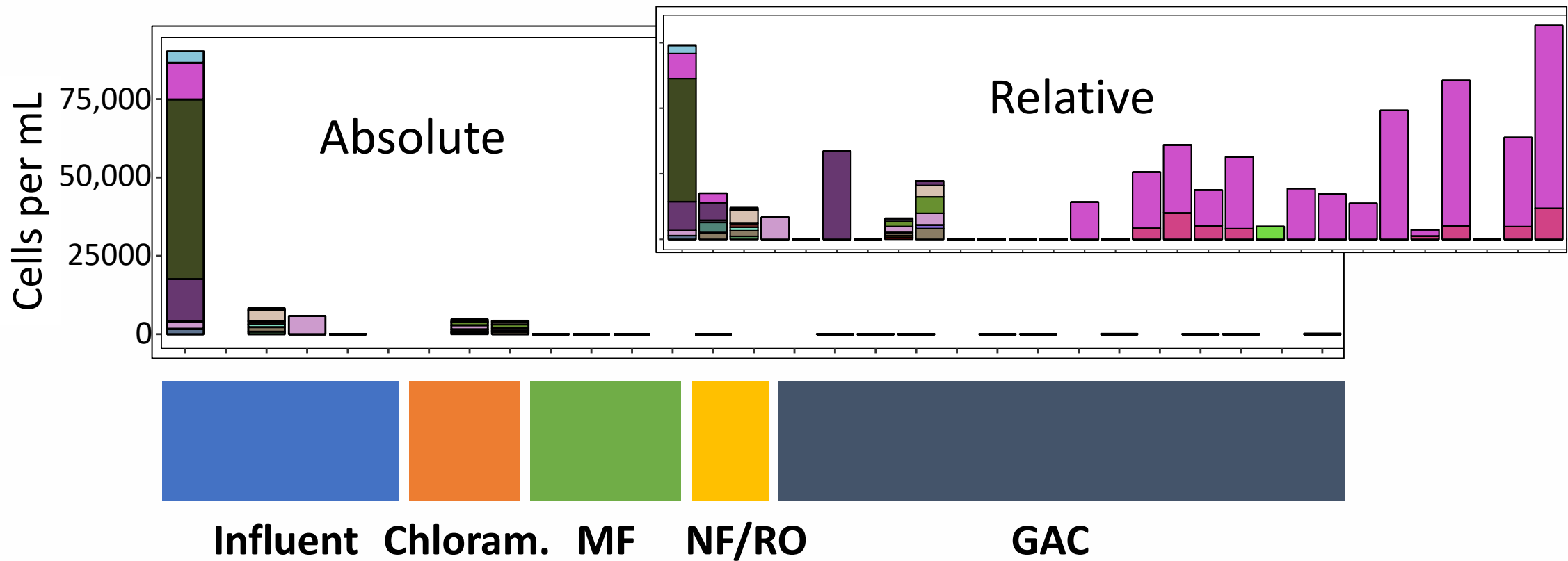
## Limitation 2: Specificity

- Most data analysis programs don't determine to species-level
- 16S amplicon sequencing is usually too short to distinguish **strains**
- Recommend qPCR (specific for pathogen)

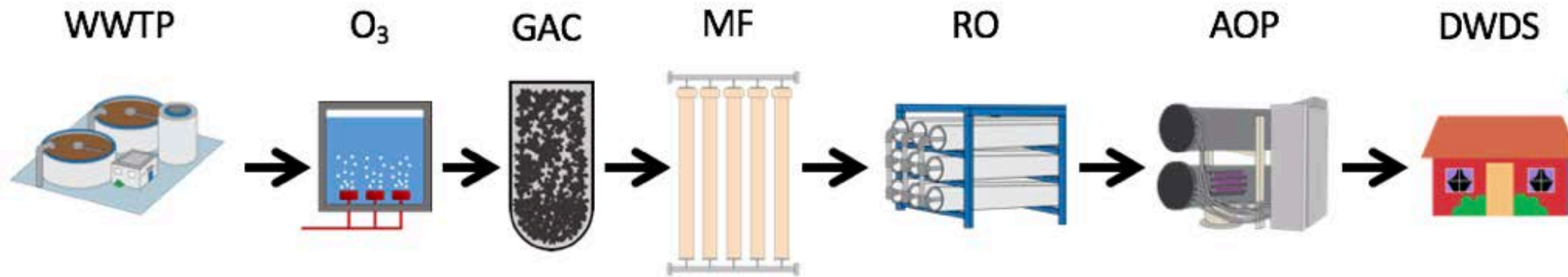
# Legionella (all species): relative abundance



# Legionella (all species): absolute abundance

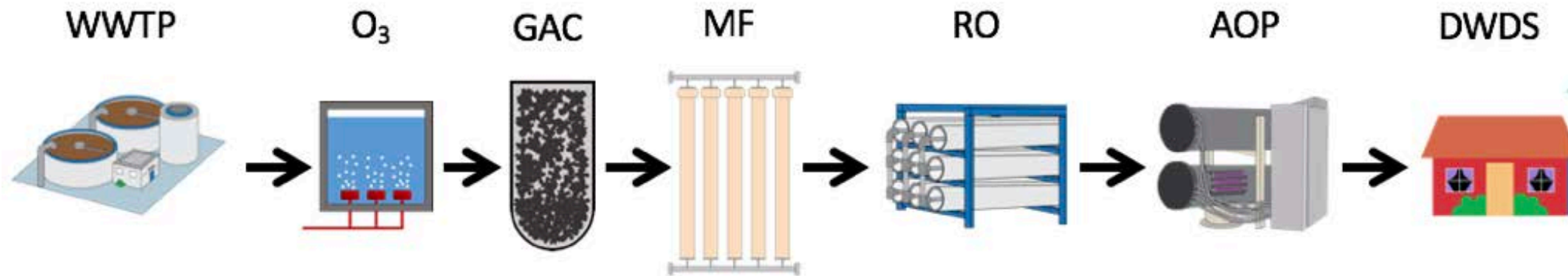


# Motivation: potable reuse microbiology



- What types of microorganisms are present and in what quantities?
  - **Predominant bacteria *change* across treatment train**

# Motivation: potable reuse microbiology



- Where might the bacteria come from? (source tracking)
  - **They may start out in place, *not sourced* from upstream**
  - **They may start out too rare to detect and then survive treatment and grow**



# Molecular methods with highly purified water: Controls matter

- Want high signal:noise ratio to prevent contamination
- Negative / blank
- Positive / DNA from known microorganisms



# Take-home messages



- Amplicon sequencing: insight into patterns within a system
- Controls are critical
- Can't detect pathogens with specificity (use qPCR)
- Should also include an absolute measure of biomass and viability or growth assays
- For functional analyses, should use metagenomics

# Costs

- Amplicon sequencing: \$5-17 per sample (other groups' estimates)
  - For highly purified water, allow 25-50% repeat rate (or filter extensively)
  - **We estimated \$18 per sample**
  - Sequenced in batches of 96 or 384, depending on desired detection limit
  - Include controls in budget
  - Consider triplicate runs per sample
- qPCR: ~\$2 per reaction, but also need calibration curve & controls
  - One assay per pathogen/ARG/other gene of interest