Monitoring variants of SARS-CoV-2 in wastewater: Overview and UC Berkeley experience

Rose Kantor, Ph.D. and wastewater testing laboratory UC Berkeley May 18, 2022

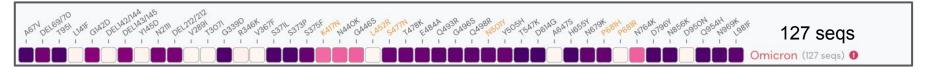
Background: challenges for variant detection in WW

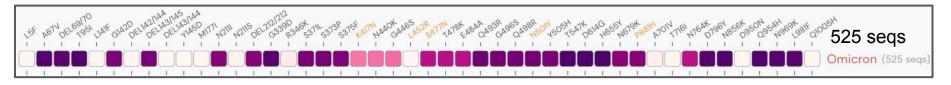
1. SARS-CoV-2 RNA is at relatively low concentrations in wastewater

- Replicate/repeat samples to account for heterogeneity & false negatives
- Concentrate virus particles
- Enrich for SARS-CoV-2 RNA (or deplete other WW components)
- Amplify RNA targets

Background: challenges for variant detection in WW

- 1. SARS-CoV-2 RNA is at relatively low concentrations in wastewater
- 2. Changing variant definitions and new variants





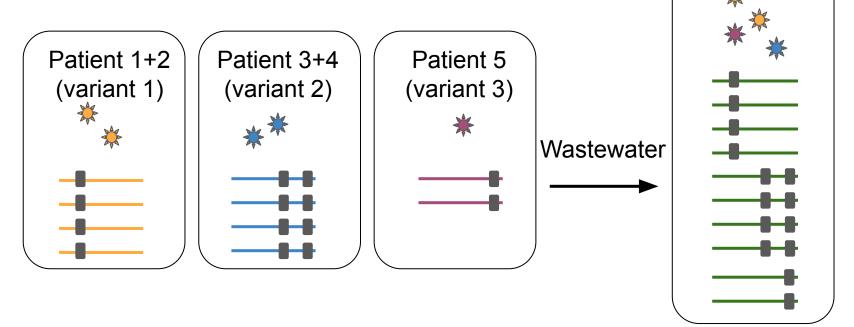


Mutation prevalence in lineage	
0	100%

Screenshots from outbreak.info

Background: challenges for variant detection in WW

- 1. SARS-CoV-2 RNA is at relatively low concentrations in wastewater
- 2. Changing variant definitions and new variants
- 3. Wastewater is a mixed strain sample



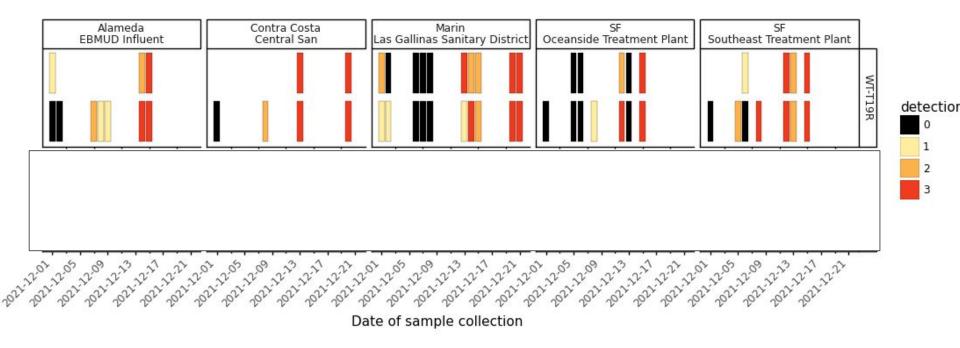
Methods choice and interpretation considerations

Consideration	PCR-based	Sequencing-based
Sensitivity		
Novel vs. known/targeted		
Quantification		
Time-to-results		
Development and reagent lead time w/ new variant		
Cost per sample		

Methods choice and interpretation considerations

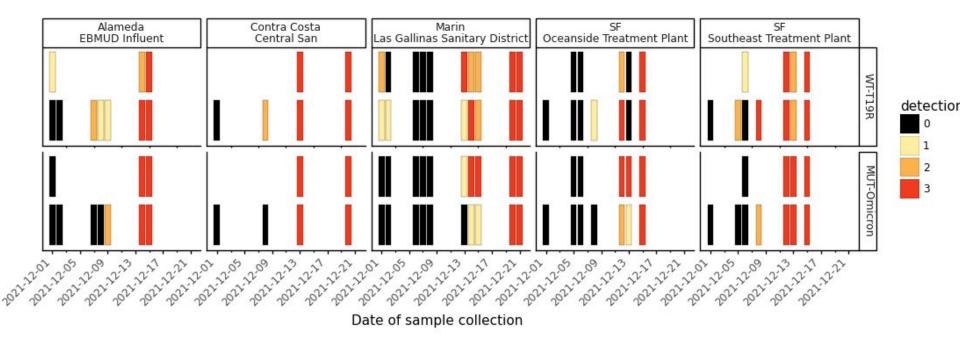
Consideration	PCR-based	Sequencing-based
Sensitivity	Higher	Lower
Novel vs. known/targeted	Known (targeted) only	Novel and known
Quantification	Absolute	Relative
Time-to-results	Faster	Slower
Development and reagent lead time w/ new variant	Longer	Shorter
Cost per sample	Lower	Higher

Results from RT-qPCR: early Omicron detection



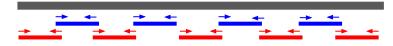
WT-T19 is a non-Delta assay

Results from RT-qPCR: early Omicron detection



Methods: Tiled Amplicon Whole Genome Sequencing

Primers designed to tile across the SARS-CoV-2 genome



Primer sets

- ARTIC
- Swift
- "Midnight"

Sequencing platforms:

- Illumina
- Nanopore

Results: Tiled Amplicon Whole Genome Sequencing

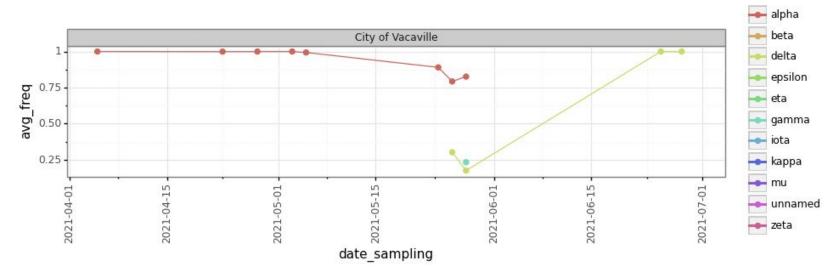
Primers designed to tile across the SARS-CoV-2 genome

Primer sets

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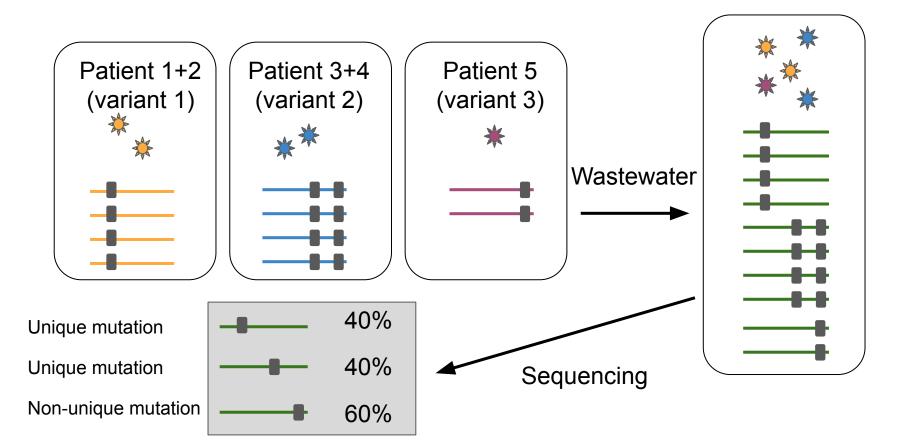
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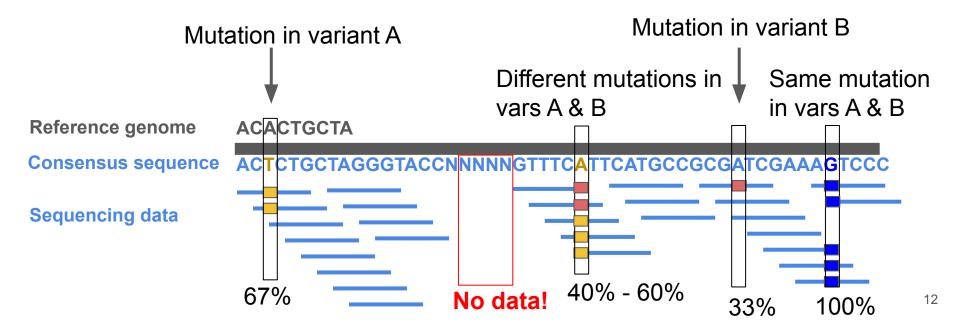


Targeted amplicon sequencing reveals linked mutations



Caveats of whole genome sequencing

- 1. Mutations are unlinked in analysis and wastewater is a mixed sample
- 2. Genomes are not always complete



Methods: Targeted Amplicon Sequencing

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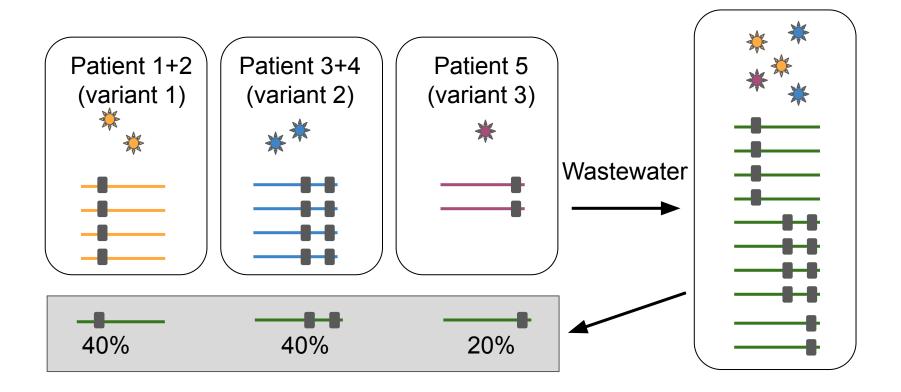
Targeted S-gene regions:

- Receptor binding domain
- N-terminal domain
- S1S2 domain

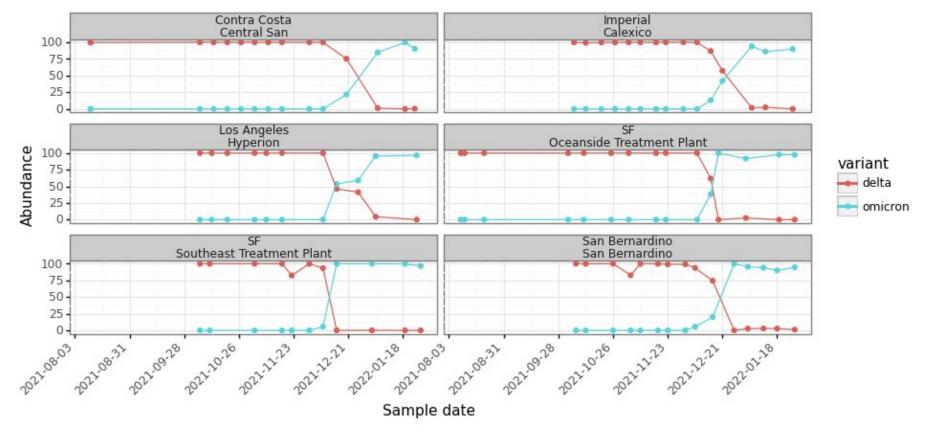
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Targeted amplicon sequencing reveals linked mutations



Results: Targeted Amplicon Sequencing



Cryptic lineages found in targeted sequencing data

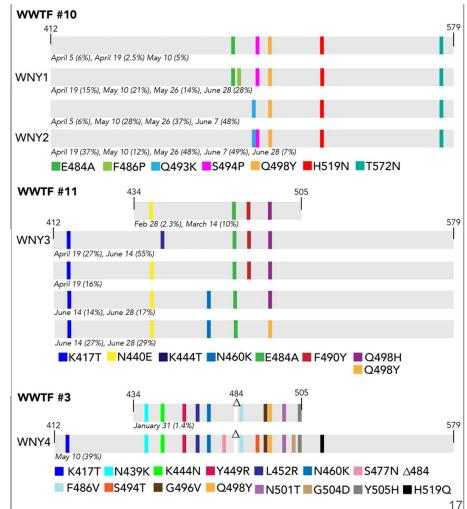
In New York City Sewage, a Mysterious Coronavirus Signal

For the past year, scientists have been looking for the source of strange coronavirus sequences that have appeared in the city's wastewater.

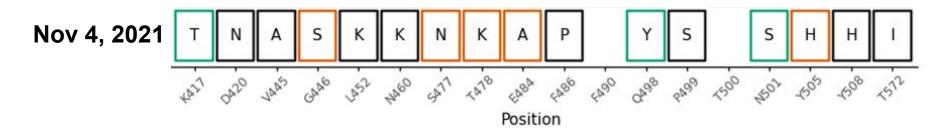
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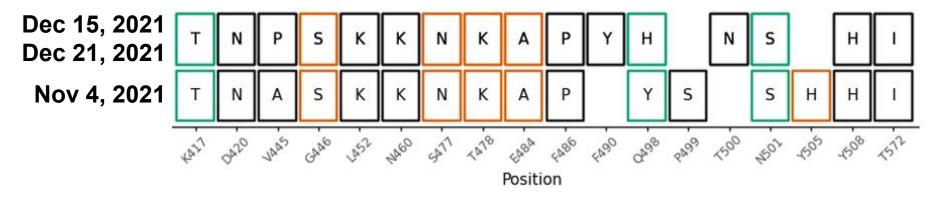
Cryptic lineages found in targeted sequencing data, CA





Gregory, Johnson, Kantor, Dennehy et al. in prep 18

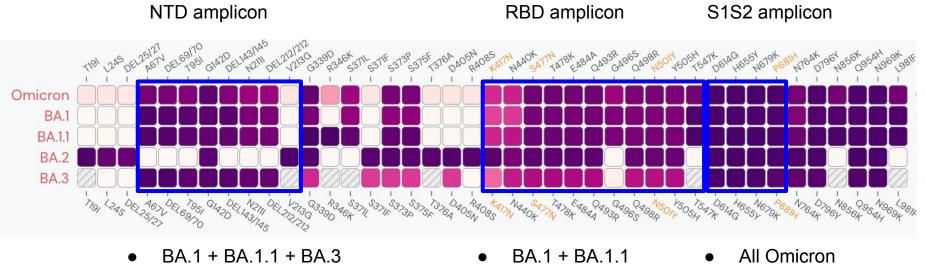
Cryptic lineages found in targeted sequencing data, CA





Gregory, Johnson, Kantor, Dennehy et al. in prep 19

Caveats of targeted amplicon sequencing: Sometimes cannot distinguish sublineages



• BA.2

• BA.2 + BA.3

Variant monitoring: barriers and directions

- Low concentrations in wastewater: sequencing may fail between surges
- Sequence data interpretation as paired with clinical data?
 - Where are there discrepancies?
 - Novel variants or sublineages?
 - Monitor variant sweeps
- Consolidated state-level dashboard?
- Fit-for-purpose monitoring effort:
 - Define the goal (e.g. detect novel variants OR track known variants discovered in clinical data)
 - Choose appropriate method
 - Analyze all data with consistent definitions of variant lineages
 - \circ Visualize, communicate within PH \rightarrow action? (e.g. decide use of monoclonal antibody treatments)

Extra slides

High-throughput sequencing of 3 s-gene regions yields exact amplicon sequences within these regions



Primers and approach from Gregory et al. 2021

Methods: PCR-based assays

Allele-specific qPCR

